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(54) Title: NOVEL LIPOSOMAL DRUG DELIVERY SYSTEMS

(57) Abstract

Drug delivery systems comprising gas filled liposomes prepared using vacuum drying gas instillation methods and having encapsulated therein a drug, and gas filled liposomes substantially devoid of liquid in the interior thereof and having encapsulated therein a drug, are described. Methods of and apparatus for preparing such liposomes and methods for employing such liposomes in drug delivery applications are also disclosed.

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NOVEL LIPOSOMAL DRUG DELIVERY SYSTEMS

RELATED APPLICATION

This application is a continuation-in-part of copending application U.S. Serial No. 569,828, filed August 20, 1990, which in turn is a continuation-in-part of application U.S. Serial No. 455,707, filed December 22, 1989, the disclosures of each of which are hereby incorporated herein by reference in their entirety.

BACKGROUND OF THE INVENTION

Field of the Invention

This invention relates to the field of drug delivery and more specifically, to gas filled liposomes prepared using vacuum drying gas instillation methods, and to gas filled liposomes substantially devoid of liquid in the interior thereof, said liposomes also having encapsulated therein a drug. The invention further relates to methods of and apparatus for preparing such drug containing gas filled liposomes and to methods for employing such liposomes as drug delivery systems.

Background of the Invention

For hundreds of years man has endeavored to develop a wide variety of drugs useful in the treatment of various diseases and disorders. Lagging noticeably behind in this effort, however, has been the development of effective means to deliver such drugs to selected sites in the body. Although some achievements have been made in this area during the last decade, *see, e.g.*, R. Baker, Controlled Release of Biologically Active Agents, John Wiley & Sons (New York 1987), new and/or better drug delivery systems are needed.

Targeted delivery means are particularly important where drug toxicity is an issue. Specific drug delivery methods potentially serve to minimize toxic side effects, lower the required dosage amounts, and decrease drug costs for the patient. The present invention is directed to addressing these and/or other important needs in the area of drug delivery.

SUMMARY OF THE INVENTION

The present invention provides novel liposomal drug delivery systems.

Specifically, in one embodiment, the present invention provides drug delivery systems comprising gas filled liposomes prepared by vacuum drying gas instillation methods and having encapsulated therein a drug, such liposomes sometimes being referred to herein as drug-containing vacuum dried gas instilled liposomes.

In another embodiment, the invention is directed to drug delivery systems comprising gas filled liposomes substantially devoid of liquid in the interior thereof and having encapsulated therein a drug.

In a further embodiment, the subject invention provides methods for preparing the liposomes of the subject invention, said methods comprising: (i) placing liposomes encapsulating a drug under negative pressure;

(ii) incubating the liposomes under the negative pressure for a time sufficient to remove substantially all liquid from the liposomes; and (iii) instilling selected gas into the liposomes until ambient pressures are achieved. Methods employing the foregoing steps are referred to herein as the vacuum drying gas instillation methods for preparing drug containing liposomes.

In a still further embodiment, the invention provides apparatus for preparing the liposomes of the invention using the vacuum drying gas instillation methods, said apparatus comprising: (i) a vessel containing liposomes having encapsulated therein a drug; (ii) means for applying negative pressure to the vessel to draw liquid from the liposomes contained therein; (iii) a conduit connecting the negative pressurizing means to the vessel, the conduit directing the flow of said liquid; and (iv) means for introducing a gas into the liposomes in the vessel.

Finally, the invention contemplates methods for the controlled delivery of drugs to a region of a patient comprising: (i) administering to the patient the gas filled liposomes prepared by vacuum drying gas instillation methods and having encapsulated therein a drug, and/or gas filled liposomes substantially devoid of liquid in the interior thereof and having encapsulated therein a drug; (ii) monitoring the liposomes using ultrasound to determine the presence of the liposomes in the region; and (iii) rupturing the liposomes using ultrasound to release the drugs in the region.

Surprisingly, the drug containing gas filled liposomes prepared by the vacuum drying gas instillation method, and the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof which may be prepared in accordance with the vacuum drying gas instillation method, possess a number of unexpected, but highly beneficial, characteristics. The liposomes of the invention exhibit intense echogenicity on ultrasound,

allowing them to be monitored effectively *in vivo*. The liposomes of the invention are also susceptible to rupture upon application of ultrasound at the peak resonant frequency of the liposomes. Such characteristics permit the subject liposomes to be employed in the novel and unexpected applications of the present invention. Further, the subject liposomes are surprisingly highly stable to pressure and/or possess a long storage life, either when stored dry or suspended in a liquid medium. Also unexpected is the ability of the liposomes during the vacuum drying gas instillation process to fill with gas and resume their original circular shape, rather than irreversibly collapse into a cup-like shape.

These and other features of the invention and the advantages thereof will be set forth in greater detail in the figures and the description below.

BRIEF DESCRIPTION OF THE FIGURES

FIGURE 1 shows an apparatus according to the present invention for preparing the drug containing vacuum dried gas instilled liposomes, and the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof prepared by the vacuum drying gas instillation method. **FIGURE 2** is a graphical representation of the dB reflectivity of gas filled liposomes substantially devoid of liquid in the interior thereof prepared by the vacuum drying gas instillation method, without any drugs encapsulated therein. The data was obtained by scanning with a 7.5 megahertz transducer using an Acoustic Imaging™ Model 5200 scanner (Acoustic Imaging, Phoenix, Arizona), and was generated by using the system test software to measure reflectivity. The system was standardized prior to each experiment with a phantom of known acoustic impedance.

DETAILED DESCRIPTION OF THE INVENTION

The present invention is directed to ultrasound contrast agents comprising gas filled liposomes prepared by vacuum drying gas instillation methods and having encapsulated therein a drug (that is, drug containing), such liposomes sometimes being referred to herein as drug containing vacuum dried gas instilled liposomes. The present invention is further directed to contrast agents comprising drug containing gas filled liposomes substantially devoid of liquid in the interior thereof.

The vacuum drying gas instillation method employed to prepare both the subject gas filled liposomes prepared by the vacuum drying gas instillation method, and the gas filled liposomes substantially devoid of liquid in the interior thereof, contemplates the following process. First, in accordance with the process, the drug containing liposomes are placed under negative pressure (that is, reduced pressure or vacuum conditions). Next, the liposomes are incubated under that negative pressure for a time sufficient to remove substantially all liquid from the liposomes, thereby resulting in substantially dried liposomes. By removal of substantially all liquid, and by substantially dried liposomes, as those phrases are used herein, it is meant that the liposomes are at least about 90% devoid of liquid, preferably at least about 95% devoid of liquid, most preferably about 100% devoid of liquid. Although the liquid is removed, the drug, with its higher molecular weight, remains behind, encapsulated in the liposome. Finally, the liposomes are instilled with selected gas by applying the gas to the liposomes until ambient pressures are achieved, thus resulting in the subject drug containing vacuum dried gas instilled liposomes of the present invention, and the drug containing gas filled liposomes of the invention substantially devoid of liquid in the interior thereof. By substantially devoid of liquid in the interior thereof, as used herein, it is meant liposomes having an interior that is at least about 90% devoid of liquid, preferably at

least about 95% devoid of liquid, most preferably about 100% devoid of liquid.

Unexpectedly, the drug containing liposomes prepared in accordance with the vacuum dried gas instillation method, and the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof, possess a number of surprising yet highly beneficial characteristics. The liposomes of the invention exhibit intense echogenicity on ultrasound, will rupture on application of peak resonant frequency ultrasound (as well as other resonant freguencies of sufficient intensity and duration), are highly stable to pressure, and/or generally possess a long storage life, either when stored dry or suspended in a liquid medium.

The ecogenicity of the liposomes and the ability to rupture the liposomes at the peak resonant frequency using ultrasound permits the controlled delivery of drugs to a region of a patient by allowing the monitoring of the liposomes following administration to a patient to determine the presence of liposomes in a desired region, and the rupturing of the liposomes using ultrasound to release the drugs in the region. Preferably, the liposomes of the invention possess a reflectivity of greater than 2 dB, preferably between about 4 dB and about 20 dB. Within these ranges, the highest reflectivity for the liposomes of the invention is exhibited by the larger liposomes, by higher concentrations of liposomes, and/or when higher ultrasound frequencies are employed. Preferably, the liposomes of the invention have a peak resonant frequency of between about 0.5 mHz and about 10 mHz. Of course, the peak resonant frequency of the gas filled liposomes of the invention will vary depending on the diameter and, to some extent, the elasticity of the liposomes, with the larger and more elastic liposomes having a lower resonant frequency than the smaller and more elastic liposomes.

The stability of the liposomes of the invention is also of great practical importance. The subject liposomes tend to have greater stability during storage than other gas filled liposomes produced via known procedures such as pressurization or other techniques. At 72 hours after formation, for example, conventionally prepared gas containing liposomes often are essentially devoid of gas, the gas having diffused out of the liposomes and/or the liposomes having ruptured and/or fused, resulting in a concomitant loss in reflectivity. In comparison, drug containing gas filled liposomes of the present invention generally have a shelf life stability of greater than about three weeks, preferably a shelf life stability of greater than about four weeks, more preferably a shelf life stability of greater than about five weeks, even more preferably a shelf life stability of greater than about three months, and often a shelf life stability that is even much longer, such as over six months, twelve months, or even two years.

Also unexpected is the ability of the liposomes during the vacuum drying gas instillation process to fill with gas and resume their original circular shape, rather than collapse into a cup-shaped structure, as the prior art would cause one to expect. *See, e.g.*, Crowe et al., Archives of Biochemistry and Biophysics, Vol. 242, pp. 240-247 (1985); Crowe et al., Archives of Biochemistry and Biophysics, Vol. 220, pp. 477-484 (1983); Fukuda et al., J. Am. Chem. Soc., Vol. 108, pp. 2321-2327 (1986); Regen et al., J. Am. Chem. Soc., Vol. 102, pp. 6638-6640 (1980).

The drug containing liposomes subjected to the vacuum drying gas instillation method of the invention may be prepared using any one of a variety of conventional liposome preparatory techniques which will be apparent to those skilled in the art. These techniques include freeze-thaw, as well as t chniques such as sonication, chelate dialysis, homogenization, solvent infusion,

microemulsification, spontaneous formation, solvent vaporization, French pressure cell technique, controlled detergent dialysis, and others, each involving preparing the liposomes in various fashions in a solution containing the desired drug so that the drug is encapsulated in the resultant liposome. Alternatively, drugs may be loaded into the liposomes using pH gradient techniques which, as those skilled in the art will recognize, is particularly applicable to drugs which either protinate or deprotinate at a particular pH. *See, e.g.*, Madden et al., Chemistry and Physics of Lipids, Vol. 53, pp. 37-46 (1990), the disclosures of which are hereby incorporated herein by reference in their entirety. The size of the drug containing liposomes can be adjusted, if desired, prior to vacuum drying and gas instillation, by a variety of procedures including extrusion, filtration, sonication, homogenization, employing a laminar stream of a core of liquid introduced into an immiscible sheath of liquid, and similar methods, in order to modulate resultant liposomal biodistribution and clearance. Extrusion under pressure through pores of defined size is, however, the preferred means of adjusting the size of the liposomes. The foregoing techniques, as well as others, are discussed, for example, in U.S. Patent No. 4,728,578; U.K. Patent Application GB 2193095 A; U.S. Patent No. 4,728,575; U.S. Patent No. 4,737,323; International Application PCT/US85/01161; Mayer et al., Biochimica et Biophysica Acta, Vol. 858, pp. 161-168 (1986); Hope et al., Biochimica et Biophysica Acta, Vol. 812, pp. 55-65 (1985); U.S. Patent No. 4,533,254; Mayhew et al., Methods in Enzymology, Vol. 149, pp. 64-77 (1987); Mayhew et al., Biochimica et Biophysica Acta, Vol 755, pp. 169-74 (1984); Cheng et al, Investigative Radiology, Vol. 22, pp. 47-55 (1987); PCT/US89/05040, U.S. Patent No. 4,162,282; U.S. Patent No. 4,310,505; U.S. Patent No. 4,921,706; and Liposome Technology, Gregoriadis, G., ed., Vol. I, pp. 29-31, 51-67 and 79-108 (CRC Press Inc., Boca Raton, FL

1984). The disclosures of each of the foregoing patents, publications and patent applications are incorporated by reference herein, in their entirety. Although any of a number of varying techniques can be employed, preferably the drug containing liposomes are prepared via microemulsification techniques. The liposomes produced by the various conventional procedures can then be employed in the vacuum drying gas instillation method of the present invention, to produce the drug containing liposomes of the present invention.

The materials which may be utilized in preparing liposomes to be employed in the vacuum drying gas instillation method of the present invention include any of the materials or combinations thereof known to those skilled in the art as suitable for liposome construction. The lipids used may be of either natural or synthetic origin. Such materials include, but are not limited to, lipids such as fatty acids, lysolipids, dipalmitoylphosphatidylcholine, phosphatidylcholine, phosphatidic acid, sphingomyelin, cholesterol, cholesterol hemisuccinate, tocopherol hemisuccinate, phosphatidylethanolamine, phosphatidyl-inositol, lysolipids, sphingomyelin, glycosphingolipids, glucolipids, glycolipids, sulphatides, lipids with ether and ester-linked fatty acids, polymerized lipids, diacetyl phosphate, stearylamine, distearoylphosphatidylcholine, phosphatidylserine, sphingomyelin, cardiolipin, phospholipids with short chain fatty acids of 6-8 carbons in length, synthetic phospholipids with asymmetric acyl chains (e.g., with one acyl chain of 6 carbons and another acyl chain of 12 carbons), 6-(5-cholesten-3β-yloxy)-1-thio-β-D-galactopyranoside, digalactosyldiglyceride, 6-(5-cholesten-3β-yloxy)hexyl-6-amino-6-deoxy-1-thio-β-D-galactopyranoside, 6-(5-cholesten-3β-yloxy)hexyl-6-amino-6-deoxyl-1-thio-α-D-mannopyranoside, dibehenoylphosphatidylcholine, dimyristoylphosphatidylcholine,

dilauroylphosphatidylcholine, and dioleoylphosphatidylcholine, and/or combinations thereof. Other useful lipids or combinations thereof apparent to those skilled in the art which are in keeping with the spirit of the present invention are also encompassed by the present invention. For example, carbohydrates bearing lipids may be employed for *in vivo* targeting, as described in U.S. Patent No. 4,310,505. Of particular interest for use in the present invention are lipids which are in the gel state (as compared with the liquid crystalline state) at the temperature at which the vacuum drying gas instillation is performed. The phase transition temperatures of various lipids will be readily apparent to those skilled in the art and are described, for example, in Liposome Technology, Gregoriadis, G., ed., Vol. I, pp. 1-18 (CRC Press, Inc. Boca Raton, FL 1984), the disclosures of which are incorporated herein by reference in their entirety. In addition, it has been found that the incorporation of at least a small amount of negatively charged lipid into any liposome membrane, although not required, is beneficial to providing highly stable liposomes. By at least a small amount, it is meant about 1 mole percent of the total lipid. Suitable negatively charged lipids will be readily apparent to those skilled in the art, and include, for example phosphatidylserine and fatty acids. Most preferred for the combined reasons of ultimate ability to rupture on application of resonant frequency ultrasound, ecogenicity and stability following the vacuum drying gas instillation process are liposomes prepared from dipalmitoylphosphatidylcholine.

Any of a variety of drugs may be encapsulated in the liposomes. By drugs, as used herein, it is meant any agent having beneficial and/or therapeutic effect on the patient. Suitable drugs include, but are not limited to: antineoplastic agents, such as platinum compounds (e.g., spiroplatin, cisplatin, and carboplatin), methotrexate, adriamycin, mitomycin, ansamitocin, blemoycin, cytosine

arabinosine, arabinosyl, anenine, mercaptopolylysine, vincristine,
busulfan, chlorambucil, melphasan (e.g., PAM, L-PAM or phenylalanine mustard), mercaptopurine, mitotane, procarbazine hydrochloride dacinomycin (actinomycin D), daunorubicin hydrochloride, doxorubicin hydrochloride, mitomycin, plicamycin (mithramycin), aminoglutethimide, estramustine phosphate sodium, flutamide, leuprolide acetate, megestrol acetate, tamoxifen citrate, testolactone, trilostane, amsacrine (m-AMSA), asparaginase (L-asparaginase) *Erwina* asparaginase, etoposide (VP-16), interferon α-2a, interferon α-2b, teniposide (VM-26), vinblastine sulfate (VLB), vincristine sulfate, bleomycin, bleomycin sulfate, methotrexate, adriamycin, cytosine arabinosine, and arabinosyl; biological response modifiers such as muramyldipeptide, muramyltripeptide, microbial cell wall components, lymphokines (e.g., bacterial endotoxin such as lipopolysaccharide, macrophage activation factor), sub-units of bacteria (such as Mycobacteria, Corynebacteria), the synthetic dipeptide N-acetyl-muramyl-L-alanyl-D-isoglutamine; genetic material such as nucleic acids, RNA, and DNA, of either natural or synthetic origin, including recombinant RNA and DNA; anti-fungal agents such as ketoconazole, nystatin, griseofulvin, flucytosone (5-fc), miconazole, amphotericin-β, ricin, and β-lactum antibiotics (e.g., sulfazecin); hormones such as growth hormone, melanocyte stimulating hormone, estradiol, beclomethasone dipropionate, betamethasone, betamethasone acetate and betamethasone sodium phosphate, vetamethasone disodium phosphate, vetamethasone sodium phosphate, cortisone acetate, dexamethasone, dexamethasone acetate, dexamethasone sodium phosphate, fluinsolide, hydrocortisone, hydrocortisone acetate, hydrocortisone cypionate, hydrocortisone sodium phosphate, hydrocortisone sodium succinate, methylprednisolone, methylprednisolone acetate, methylprednisolone sodium succinate,

paramethasone acetate, prednisolone, prednisolone acetate, prednisolone sodium phosphate, prednisolone tebutate, prednisone, triamcinolone, triamcinolone acetonide, triamcinolone diacetate, triamcinolone hexacetonide and fludrocortisone acetate; vitamins such as cyanocobalamin and α-tocopherol; peptides, such as manganese super oxide dimutase; enzymes such as alkaline phosphatase; anti-allergic agents such as amelexanox; anti-coagulation agents such as phenprocoumon and heparin; circulatory drugs such as propranolol; metabolic potentiators such as glutathione; antituberculars such as para-aminosalicylic acid, isoniazid capreomycin sulfate cycloserine, ethambutol hydrochloride ethionamide, pyrazinamide, rifampin, and streptomycin sulfate; antivirals such as acyclovir, amantadine azidothymidine (AZT or Zidovudine), ribavirin and vidarabine monohydrate (adenine arabinoside, ara-A); antianginals such as diltiazem, nifedipine, verapamil, erythrityl tetranitrate, isosorbide dinitrate, nitroglycerin (glyceryl trinitrate) and pentaerythritol tetranitrate; anticoagulants such as phenprocoumon, heparin; antibiotics such as dapsone, chloramphenicol, neomycin, cefaclor, cefadroxil, cephalexin, cephradine erythromycin, clindamycin lincomycin, amoxicillin, ampicillin, bacampicillin, carbenicillin, cloxacillin, cyclacillin, picloxacillin, hetacillin, methicillin, nafcillin, oxacillin, penicillin G, penicillin V, ticarcillin fifampin and tetracycline; antiinflammatories such as difunisal, ibuprofen, indomethacin, meclofenamate, mefenamic acid, naproxen, oxyphenbutazone, phenylbutazone, piroxicam, culindac, tolmetin, aspirin and salicylates; antiprotozoans such as chloroquine, hydroxycloroquine, metronidazole, quinine and meglemine antimonate; antirheumatics such as penicillamine; narcotics such as paregoric; opiates such as codeine, heroin, methadone, morphine and opium; cardiac glycosides such as deslanoside, digitoxin, digoxin, digitalin and digitalis; neuromuscular blockers such as atracurium besylate,

gallamine triethiodide, hexafluorenium bromide, metocurine iodide, pancuronium bromide, succinylcholine chloride (suxamethonium chloride), tubocurarine chloride and vecuronium bromide; sedatives (hypnotics) such as amobarbital, amobarbital sodium, aprobarbital, butabarbital sodium, chloral hydrate, ethchlorvynol, ethinamate, flurazepam hydrochloride, glutethimide, methotrimeprazine hydrochloride, methyprylon, midazolam hydrochloride, paraldehyde, pentobarbital, pentobarbital sodium, phenobarbital sodium, secobarbital sodium, talbutal, temazepam and triazolam; local anesthetics such as bupivacaine hydrochloride, chloroprocaine hydrochloride, etidocaine hydrochloride, lidocaine hydrochloride, mepivacaine hydrochloride, procaine hydrochloride and tetracaine hydrochloride; general anesthetics such as droperidol, etomidate, fentanyl citrate with droperidol, ketamine hydrochloride, methohexital sodium and thiopental sodium; and radioactive particles or ions such as strontium, iodide rhenium and yttrium.

Similarly, prodrugs may be encapsulated in the liposomes, and are included within the ambit of the terms drug or drugs, as used herein. Prodrugs are well known in the art, and include, inactive drug precursors which, when exposed to high temperature, cavitation and/or pressure, in the presence of oxygen or otherwise, or when released from the liposomes, will form active drugs. Such prodrugs can be activated in the method of the invention, upon the application of ultrasound to the prodrug-containing liposomes with the resultant cavitation, heating, pressure, and/or release from the liposomes. Suitable prodrugs will be apparent to those skilled in the art, and are described, for example, in Sinkula et al., J. Pharm. Sci., Vol. 64, pp. 181-210 (1975), the disclosures of which are hereby incorporated herein by reference in their entirety. Prodrugs, for example, may comprise inactive forms of the active drugs wherein a chemical group is

present on the prodrug which renders it inactive and/or confers solubility or some other property to the drug. In this form, the prodrugs are generally inactive, but once the chemical group has been cleaved from the prodrug, by heat, cavitation, pressure, and/or by enzymes in the surrounding environment or otherwise, the active drug is generated. Such prodrugs are well described in the art, and comprise a wide variety of drugs bound to chemical groups through bonds such as esters to short, medium or long chain aliphatic carbonates, hemiesters of organic phosphate, pyrophosphate, sulfate, amides, amino acids, azo bonds, carbamate, phosphamide, glucosiduronate, N-acetylglucosaminide and β-glucoside. Examples of drugs with the parent molecule and the reversible modification or linkage are as follows: convallatoxin with ketals, hydantoin with alkyl esters, chlorphenesin with glycine or alanine esters, acetaminophen with caffeine complex, acetylsalicylic acid with THAM salt, acetylsalicylic acid with acetamidophenyl ester, naloxone with sulfate ester, 15-methylprostaglandin $F_{2\alpha}$ with methyl ester, procaine with polyethylene glycol, erythromycin with alkyl esters, clindamycin with alkyl esters or phosphate esters, tetracycline with betaine salts, 7-acylaminocephalosporins with ring-substituted acyloxybenzyl esters, nandrolone with phenylproprionate decanoate esters, estradiol with enol ether acetal, methylprednisolone with acetate esters, testosterone with n-acetylglucosaminide glucosiduronate (trimethylsilyl) ether, cortisol or prednisolone or dexamethasone with 21-phosphate esters. Prodrugs may also be designed as reversible drug derivatives and utilized as modifiers to enhance drug transport to site-specific tissues. Examples of parent molecules with reversible modifications or linkages to influence transport to a site specific tissue and for enhanced therapeutic effect include isocyanate with haloalkyl nitrosurea, testosterone with propionate ester, methotrexate (3-5'-dichloromethotrexate) with dialkyl esters, cytosine

arabinoside with 5'-acylate, nitrogen mustard (2,2'-dichloro-N-methyldiethylamine), nitrogen mustard with aminomethyl tetracycline, nitrogen mustard with cholesterol or estradiol or dehydroepiandrosterone esters and nitrogen mustard with azobenzene. As one skilled in the art would recognize, a particular chemical group to modify a given drug may be selected to influence the partitioning of the drug into either the membrane or the internal space of the liposomes. The bond selected to link the chemical group to the drug may be selected to have the desired rate of metabolism, e.g., hydrolysis in the case of ester bonds in the presence of serum esterases after release from the gas filled liposomes. Additionally, the particular chemical group may be selected to influence the biodistribution of the drug employed in the gas filled drug carrying liposome invention, e.g., N,N-bis(2-chloroethyl)-phosphorodiamidic acid with cyclic phosphoramide for ovarian adenocarcinoma. Additionally, the prodrugs employed within the gas filled liposomes may be designed to contain reversible derivatives which are utilized as modifiers of duration of activity to provide, prolong or depot action effects. For example, nicotinic acid may be modified with dextran and carboxymethlydextran esters, streptomycin with alginic acid salt, dihydrostreptomycin with pamoate salt, cytarabine (ara-C) with 5'-adamantoate ester, ara-adenosine (ara-A) with 5-palmitate and 5'-benzoate esters, ampho- tericin-β with methyl esters, testosterone with 17-β-alkyl esters, estradiol with formate ester, prostaglandin with 2-(4-imidazolyl)ethylamine salt, dopamine with amino acid amides, chloramphenicol with mono- and bis(trimethylsilyl) ethers, and cycloguanil with pamoate salt. In this form, a depot or reservoir of long-acting drug may be released *in vivo* from the gas filled prodrug bearing liposomes. In addition, compounds which are generally thermally labile may be utilized to create toxic fee radical compounds. Compounds with azolinkages,

peroxides and disulfide linkages which decompose with high temperature are preferred. With this form of prodrug, azo, peroxide or disulfide bond containing compounds are activated by cavitation and/or increased heating caused by the interaction of high energy sound with the gas filled liposomes to create cascades of free radicals from these prodrugs entrapped therein. A wide variety of drugs or chemicals may constitute these prodrugs, such as azo compounds, the general structure of such compounds being R-N=N-R, wherein R is a hydrocarbon chain, where the double bond between the two nitrogen atoms may react to create free radical products *in vivo*. Exemplary drugs or compounds which may be used to create free radical products include azo containing compounds such as azobenzene, 2,2'-azobisisobutyronitrile, azodicarbonamide, azolitmin, azomycin, azosemide, azosulfamide, azoxybenzene, aztreonam, sudan III, sulfachrysoidine, sulfamidocnrysoidine and sulfasalazine, compounds containing disulfide bonds such as sulbentine, thiamine disulfide, thiolutin, thiram, compounds containing peroxides such as hydrogen peroxide and benzoylperoxide, 2,2'-azobis(2-amidopropane) dihydrochloride, and 2,2'-azobis(2,4-dimethylvaleronitrile). A gas filled liposome filled with oxygen gas should create extensive free radicals with cavitation. Also, metal ions from the transition series, especially manganese, iron and copper can increase the rate of formation of reactive oxygen intermediates from oxygen. By encapsulating metal ions within the liposomes, the formation of free radicals *in vivo* can be increased. These metal ions may be incorporated into the liposomes as free salts, as complexes, e.g., with EDTA, DTPA, DOTA or desferrioxamine, or as oxides of the metal ions. Additionally, derivatized complexes of the metal ions may be bound to the lipid head groups, or lipophilic complexes of the ions may be incorporated into the lipid bilayer. When exposed to thermal stimulation, e.g., cavitation, these metal ions

then will increase the rate of formation of reactive oxygen intermediates. Further, radiosensitizers such as metronidazole and misonidazole may be incorporated into the gas filled liposomes to create free radicals on thermal stimulation.

By way of an example of the use of prodrugs, an acylated chemical group may be bound to a drug via an ester linkage which would readily cleave *in vivo* by enzymatic action in serum. The acylated prodrug is incorporated into the gas filled liposome of the invention. The liposomes may also be designed so that there is a symmetric or an asymmetric distribution of the drug both inside and outside of the liposome. When the gas filled liposome is popped by the sonic pulse from the ultrasound, the prodrug encapsulated by the liposome will then be exposed to the serum. The ester linkage is then cleaved by esterases in the serum, thereby generating the drug.

Similarly, ultrasound may be utilized not only to pop the gas filled liposome, but also to cause thermal effects which may increase the rate of the chemical cleavage and the release of the active from the prodrug.

As one skilled in the art will recognize, the particular chemical structure of the drugs may be selected or modified to achieve desired solubility such that the drug may either be encapsulated within the internal aqueous space of the liposome or into the lipid membrane. The membrane bound drug may bear one or more acyl chains such that, when the bubble is popped or heated or via cavitation, the acylated drug may then leave the membrane and/or the drug may be cleaved from the acyl chains chemical group. Similarly, other drugs may be formulated with a hydrophobic group which is aromatic or sterol in structure to incorporate into the membrane.

To prepare the drug containing liposomes, and by way of general guidance, dipalmitoylphosphatidylcholine liposomes, for example,may be prepared by suspending

dipalmitoyl- phosphatidylcholine lipids in phosphate buffered saline or water containing the drug to be encapsulated, and heating the lipids to about 50°C, a temperature which is slightly above the 45°C temperature required for transition of the dipalmitoylphosphatidylcholine lipids from a gel state to a liquid crystalline state, to form drug containing liposomes. To prepare multilamellar vesicles of a rather heterogeneous size distribution of around 2 microns, the liposomes may then be mixed gently by hand while keeping the liposome solution at a temperature of about 50°C. The temperature is then lowered to room temperature, and the liposomes remain intact. Extrusion of dipalmitoylphosphatidylcholine liposomes through polycarbonate filters of defined size may, if desired, be employed to make liposomes of a more homogeneous size distribution. A device useful for this technique is an extruder device (Extruder Device™, Lipex Biomembranes, Vancouver, Canada) equipped with a thermal barrel so that extrusion may be conveniently accomplished above the gel state-liquid crystalline transition temperature for lipids.

For lipophilic drugs which are sparingly soluble in aqueous media, such drugs may be mixed with the lipids themselves prior to forming the liposomes. For example, amphotericin may be suspended with the dried lipids (e.g., 8:2 molar ratio of egg phosphatidylcholine and cholesterol in chloroform and mixed with the lipids). The chloroform is then evaporated (note that other suitable organic solvents may also be used, such as ethanol or ether) and the dried lipids containing a mixture of the lipophilic drug are then resuspended in aqueous media, e.g., sterile water or physiologic saline. This process may be used for a variety of lipophilic drugs such as corticosteroids to incorporate lipophilic drugs into the liposome membranes. The resulting liposomes are then dried, subjected to the vacuum gas instillation method as described above.

Alternatively, and again by way of general guidance, conventional freeze-thaw procedures may be used to produce either oligolamellar or unilamellar dipalmitoylphosphatidyl- choline liposomes. After the freeze-thaw procedures, extrusion procedures as described above may then be performed on the liposomes.

The drug containing liposomes thus prepared may then be subjected to the vacuum drying gas instillation process of the present invention, to produce the drug containing vacuum dried gas instilled liposomes, and the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof, of the invention. In accordance with the process of the invention, the drug containing liposomes are placed into a vessel suitable for subjecting to the liposomes to negative pressure (that is, reduced pressure or vacuum conditions). Negative pressure is then applied for a time sufficient to remove substantially all liquid from the liposomes, thereby resulting in substantially dried liposomes. As those skilled in the art would recognize, once armed with the present disclosure, various negative pressures can be employed, the important parameter being that substantially all of the liquid has been removed from the liposomes. Generally, a negative pressure of at least about 700 mm Hg and preferably in the range of between about 700 mm Hg and about 760 mm Hg (gauge pressure) applied for about 24 to about 72 hours, is sufficient to remove substantially all of the liquid from the liposomes. Other suitable pressures and time periods will be apparent to those skilled in the art, in view of the disclosures herein.

Finally, a selected gas is applied to the liposomes to instill the liposomes with gas until ambient pressures are achieved, thereby resulting in the drug containing vacuum dried gas instilled liposomes of the invention, and in the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof. Preferably, gas instillation occurs slowly, that is, over

a time period of at least about 4 hours, most preferably over a time period of between about 4 and about 8 hours. Various biocompatible gases may be employed. Such gases include air, nitrogen, carbon dioxide, oxygen, argon, xenon, neon, helium, or any and all combinations thereof. Other suitable gases will be apparent to those skilled in the art, the gas chosen being only limited by the proposed application of the liposomes.

The above described method for production of liposomes is referred to hereinafter as the vacuum drying gas instillation process.

If desired, the liposomes may be cooled, prior to subjecting the liposomes to negative pressure, and such cooling is preferred. Preferably, the liposomes are cooled to below 0°C, more preferably to between about -10°C and about -20°C, and most preferably to -10°C, prior to subjecting the liposomes to negative pressure. Upon reaching the desired negative pressure, the liposomes temperature is then preferably increased to above 0°C, more preferably to between about 10°C and about 20°C, and most preferably to 10°C, until substantially all of the liquid has been removed from the liposomes and the negative pressure is discontinued, at which time the temperature is then permitted to return to room temperature.

If the liposomes are cooled to a temperature below 0°C, it is preferable that the vacuum drying gas instillation process be carried out with liposomes either initially prepared in the presence of cryoprotectants, or liposomes to which cryoprotectants have been added prior to carrying out the vacuum drying gas instillation process of the invention. Such cryoprotectants, while not mandatorily added, assist in maintaining the integrity of liposome membranes at low temperatures, and also add to the ultimate stability of the membranes. Preferred cryoprotectants are trehalose, glycerol, polyethyleneglycol (especially polyethyleneglycol of

molecular weight 400), raffinose, sucrose and sorbitol, with trehalose being particularly preferred.

It has also been surprisingly discovered that the liposomes of the invention are highly stable to changes in pressure. Because of this characteristic, extrusion of the liposomes through filters of defined pore size following vacuum drying and gas instillation can be carried out, if desired, to create liposomes of relatively homogeneous and defined pore size.

For storage prior to use, the drug containing liposomes of the present invention may be suspended in an aqueous solution, such as a saline solution (for example, a phosphate buffered saline solution), or simply water, and stored preferably at a temperature of between about 2°C and about 10°C, preferably at about 4°C. Preferably, the water is sterile. Most preferably, the liposomes are stored in a hypertonic saline solution (e.g., about 0.3 to about 0.5% NaCl), although, if desired, the saline solution may be isotonic. The solution also may be buffered, if desired, to provide a pH range of pH 6.8 to pH 7.4. Suitable buffers include, but are not limited to, acetate, citrate, phosphate and bicarbonate. Dextrose may also be included in the suspending media. Preferably, the aqueous solution is degassed (that is, degassed under vacuum pressure) prior to suspending the liposomes therein. Bacteriostatic agents may also be included with the liposomes to prevent bacterial degradation on storage. Suitable bacteriostatic agents include but are not limited to benzalkonium chloride, benzethonium chloride, benzoic acid, benzyl alcohol, butylparaben, cetylpyridinium chloride, chlorobutanol, chlorocresol, methylparaben, phenol, potassium benzoate, potassium sorbate, sodium benzoate and sorbic acid. One or more antioxidants may further be included with the gas filled liposomes to prevent oxidation of the lipid. Suitable antioxidants include tocopherol, ascorbic acid and ascorbyl palmitate. Liposomes prepared in the various

foregoing manners may be stored for at least several weeks or months. Liposomes of the present invention may alternatively, if desired, be stored in their dried, unsuspended form, and such liposomes also have a shelf life of greater than several weeks or months. Specifically, the liposomes of the present invention, stored either way, generally have a shelf life stability of greater than about three weeks, preferably a shelf life stability of greater than about four weeks, more preferably a shelf life stability of greater than about five weeks, even more preferably a shelf life stability of greater than about three months, and often a shelf life stability that is even much longer, such as over six months, twelve months or even two years.

As another aspect of the invention, useful apparatus for preparing the drug containing vacuum dried gas instilled liposomes, and the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof, of the invention is also presented. Specifically, there is shown in Figure 1 a preferred apparatus for vacuum drying liposomes and instilling a gas into the dried liposomes. The apparatus is comprised of a vessel 8 for containing drug containing liposomes 19. If desired, the apparatus may include an ice bath 5 containing dry ice 17 surrounding the vessel 8. The ice bath 5 and dry ice 17 allow the liposomes to be cooled to below 0°C. A vacuum pump 1 is connected to the vessel 8 via a conduit 15 for applying a sustained negative pressure to the vessel. In the preferred embodiment, the pump 1 is capable of applying a negative pressure of at least about 700 mm Hg, and preferably a negative pressure in the range of about 700 mm Hg to about 760 mm Hg (gauge pressure). A manometer 6 is connected to the conduit 15 to allow monitoring of the negative pressure applied to the vessel 8.

In order to prevent liquid removed from the liposomes from entering the pump 1, a series of traps are

connected to the conduit 15 to assist in collecting the liquid (and liquid vapor, all collectively referred to herein as liquid) drawn from the liposomes. In a preferred embodiment, two traps are utilized. The first trap is preferably comprised of a flask 7 disposed in an ice bath 4 with dry ice 17. The second trap is preferably comprised of a column 3 around which tubing 16 is helically arranged. The column 3 is connected to the conduit 15 at its top end and to one end of the tubing 16 at its bottom end. The other end of the tubing 16 is connected to the conduit 15. As shown in Figure 1, an ice bath 2 with dry ice 17 surrounds the column 3 and tubing 16. If desired, dry ice 17 can be replaced with liquid nitrogen, liquid air or other cryogenic material. The ice baths 2 and 4 assist in collecting any liquid and condensing any liquid vapor drawn from the liposomes for collection in the traps. In preferred embodiments of the present invention the ice traps 2 and 4 are each maintained at a temperature of least about -70°C.

A stopcock 14 is disposed in the conduit 15 upstream of the vessel 8 to allow a selected gas to be introduced into the vessel 8 and into the liposomes 19 from gas bottle 18.

Apparatus of the present invention are utilized by placing the drug containing liposomes 19 into vessel 8. In a preferable embodiment, ice bath 5 with dry ice 17 is used to lower the temperature of the liposomes to below 0°C, more preferably to between about -10°C and about -20°C, and most preferably to -10°C With stopcocks 14 and 9 closed, vacuum pump 1 is turned on. Stopcocks 10, 11, 12 and 13 are then carefully opened to create a vacuum in vessel 8 by means of vacuum pump 1. The pressure is gauged by means of manometer 6 until negative pressure of at least about 700 mm Hg, and preferably in the range of between about 700 mm Hg and about 760 mm Hg (gauge pressure) is achieved. In preferred embodiments of the present invention vessel 7, cooled by ice bath 4 with dry

ice 17, and column 3 and coil 16, cooled by ice bath 2 with dry ice 17, together or individually condense liquid vapor and trap liquid drawn from the liposomes so as to prevent such liquids and liquid vapor from entering the vacuum pump 1. In preferred embodiments of the present invention, the temperature of ice traps 2 and 4 are each maintained at a temperature of at least about -70°C. The desired negative pressure is generally maintained for at least 24 hours as liquid and liquid vapor is removed from the liposomes 19 in vessel 8 and frozen in vessels 3 and 7. Pressure within the system is monitored using manometer 6 and is generally maintained for about 24 to about 72 hours, at which time substantially all of the liquid has been removed from the liposomes. At this point, stopcock 10 is slowly closed and vacuum pump 1 is turned off. Stopcock 14 is then opened gradually and gas is slowly introduced into the system from gas bottle 18 through stopcock 14 via conduit 15 to instill gas into the drug containing liposomes 19 in vessel 8. Preferably the gas instillation occurs slowly over a time period of at least about 4 hours, most preferably over a time period of between about 4 and about 8 hours, until the system reaches ambient pressure.

The drug containing vacuum dried gas instilled liposomes and the drug containing gas filled liposomes substantially devoid of liquid in the interior thereof, of the present invention, have superior characteristics as drug delivery vehicles. Specifically, the present invention may be employed in the controlled delivery of drugs to a region of a patient wherein the patient is administered the drug containing liposome of the present invention, the liposomes are monitored using ultrasound to determine the presence of the liposomes in the region, and the liposomes are then ruptured using ultrasound to release the drugs in the region. The patient may be any type of mammal, but is most preferably human. By region of a patient, it is meant the whole patient, or a

particular area or portion of the patient. For example, by using the method of the invention, drug delivery may be effected in a patient's heart, and a patient's vasculature (that is, venous or arterial systems). The invention is also particularly useful in delivering drugs to a patient's left heart, a region not easily reached heretofore with drug delivery. Drugs may also be easily delivered to the liver, spleen and kidney regions of a patient, as well as other regions, using the present methods.

The rupturing of the drug containing liposomes of the invention is surprisingly easily carried out by applying ultrasound of a certain frequency to the region of the patient where therapy is desired, after the liposomes have been administered to or have otherwise reached that region. Specifically, it has been unexpectedly found that when ultrasound is applied at a frequency corresponding to the peak resonant frequency of the drug containing gas filled liposomes, the liposomes will rupture and release their contents. The peak resonant frequency can be determined either *in vivo* or *in vitro*, but preferably *in vivo*, by exposing the liposomes to ultrasound, receiving the reflected resonant frequency signals and analyzing the spectrum of signals received to determine the peak, using conventional means. The peak, as so determined, corresponds to the peak resonant frequency (or second harmonic, as it is sometimes termed). The gas filled liposomes will also rupture when exposed to non-peak resonant frequency ultrasound, however, the intensity (wattage) and duration (time) must be higher in order to cause the liposomes to rupture. This higher energy results in greatly increased heating, which may not be desireable. By adjusting the frequency of the energy to match the peak resonant frequency, the efficiency of rupture and drug release is improved, appreciable tissue heating does not generally occur, (frequently no increase in temperature above about 2°C), and less overall energy

is required. Thus, application of ultrasound at the peak resonant frequency while not required is most preferred.

Any of the various types of diagnostic ultrasound imaging devices may be employed in the practive of the invention, the particular type or model of the device not being critical to the method of the invention. Also suitable are devices designed for administering ultrasonic hyperthermia, such devices being described in U.S. Patent Nos. 4,620,546, 4,658,828, and 4,586,512, the disclosures of each of which are hereby incorporated herein by reference in their entirety. Preferably, the device employs a resonant frequency (RF) spectral analyzer. Ultrasound is generally initiated at lower intensity and duration, preferably at peak resonant frequency, and then intensity, time, and/or resonant frequency increased until liposomal rupturing occurs.

Although application of the various principles will be readily apparent to one skilled in the art, once armed with the present disclosure, by way of general guidance, for gas filled liposomes of about 1.5 to about 2.0 microns diameter, the resonant frequency will generally be in the range of about 7.5 megahertz. By adjusting the focal zone to the center of the target tissue (e.g., the tumor) the gas filled liposomes can be visualized under real time ultrasound as they accumulate within the target tissue. Using the 7.5 megahertz curved array transducer as an example, adjusting the power delivered to the transducer to maximum and adjusting the focal zone within the target tissue, the spatial peak temporal average (SPTA) power will then be a maximum of approximately 5.31 mW/cm^2 in water. This power will cause some release of drug from the gas filled liposomes, but much greater release can be accomplished by using higher power. By switching the transducer to the doppler mode, higher power outputs are available, up to 2.5 watts per cm^2 from the same transducer. With the machine operating in doppler mode, the power can be delivered to a selected

focal zone within the target tissue and the gas filled liposomes can be made to release their drugs. Selecting the transducer to match the resonant frequency of the gas filled liposomes will make this process of drug release even more efficient. For larger diameter gas filled liposomes, e.g., greater than 3 microns in size, a lower frequency transducer may be more effective in accomplishing drug release. For example, a lower frequency transducer of 3.5 megahertz (20 mm curved array model) may be selected to correspond to the resonant frequency of the gas filled liposomes. Using this transducer, 101.6 milliwatts per cm^2 may be delivered to the focal spot, and switching to doppler mode will increase the power output (SPTA) to 1.02 watts per cm^2. To use the phenomenon of cavitation to release and/or activate the drugs/prodrugs within the gas filled liposomes lower frequency energies may be used, as cavitation occurs more effectively at lower frequencies. Using a 0.757 megahertz transducer driven with higher voltages (as high as 300 volts) cavitation of solutions of gas filled liposomes will occur at thresholds of about 5.2 atmospheres.

Liposomes of the present invention may be of varying sizes, but preferably are of a size range wherein they have a mean outside diameter between about 30 nanometers and about 10 microns, with the preferable mean outside diameter being about 2 microns. As is known to those skilled in the art, liposome size influences biodistribution and, therefore, different size liposomes may be selected for various purposes. For intravascular use, for example, liposome size is generally no larger than about 5 microns, and generally no smaller than about 30 nanometers, in mean outside diameter. To provide drug delivery to organs such as the liver and to allow differentiation of tumor from normal tissue, smaller liposomes, between about 30 nanometers and about 100 nanometers in mean outside diameter, are useful. With

the smaller liposomes, resonant frequency ultrasound will generally be higher than for the larger liposomes.

The gas filled drug containing liposomes of the present invention may be used with any of the various types of ultrasound imaging devices designed for administering ultrasonic hyperthermia. Such devices are well known and are described in U.S. Patent Nos. 4,620,546, 4,658,828 and 4,586,512, the disclosures of each of which are incorporated herein by reference in their entirety, and are commonly employed in physical therapy and sports medicine. It is preferable to have direct access to the resonant frequency data from the ultrasonic data using commercially available software, rather than to reformat this data (e.g., via Fourrier transform) into two dimensional or three dimensional images, to determine the peak resonant frequency. Also, the transducer probes may be external or may be implanted, if desired.

As one skilled in the art would recognize, administration of drug delivery systems of the present invention may be carried out in various fashions, such as intravascularly, intralymphatically, parenterally, subcutaneously, intramuscularly, intraperitoneally, interstitially, hyperbarically, orally, or intratumorly, using a variety of dosage forms. One preferred route of administration is intravascularly. For intravascular use, the drug delivery system is generally injected intravenously, but may be injected intraarterially as well. The useful dosage to be administered and the mode of administration will vary depending upon the age, weight, and mammal to be treated, and the particular therapeutic application intended. Typically, dosage is initiated at lower levels and increased until the desired therapeutic effect is achieved. Generally, the drug delivery systems of the invention are administered in the form of an aqueous suspension such as in water or a saline solution (e.g., phosphate buffered saline). Preferably,

the water is sterile. Also, preferably the saline solution is a hypertonic saline solution (e.g., about 0.3 to about 0.5% NaCl), although, if desired, the saline solution may be isotonic. The solution may also be buffered, if desired, to provide a pH range of pH 6.8 to pH 7.4. In addition, dextrose may be preferably included in the media. Preferably, the aqueous solution is degassed (that is, degassed under vacuum pressure) prior to suspending the liposomes therein.

The liposomes of the present invention are believed to differ from the liposomes of the prior art in a number of respects, both in physical and in functional characteristics. For example, the liposomes of the invention are substantially devoid of liquid in the interior thereof. By definition, liposomes in the prior art have been characterized by the presence of an aqueous medium. *See, e.g.*, Dorland's Illustrated Medical Dictionary, p. 946, 27th ed. (W.B. Saunders Company, Philadelphia 1988). Moreover, the present liposomes surprisingly exhibit intense ecogenicity on ultrasound, are susceptible to rupture upon application of ultrasound at the peak resonant frequency of the liposomes, and possess a long storage life, characteristics of great benefit to the use of the liposomes as drug delivery systems.

There are various other applications for liposomes of the invention, beyond those described in detail herein. Such additional uses, for example, include such applications as hyperthermia potentiators for ultrasound and as contrast agents for ultrasonic imaging. Such additional uses and other related subject matter are described and claimed in Applicant's patent applications filed concurrently herewith entitled "Method For Producing Localized Therapeutic Heat To Biological Tissues and Fluids Using Gas Filled Liposomes" and "Gas Filled Liposomes And Their Use As Ultrasonic Contrast Agents",

the disclosures of each of which are incorporated herein by reference in their entirety.

The present invention is further described in the following examples. Examples 1-10 are prophetic examples that describe the preparation, testing and use of the drug containing vacuum dried gas instilled liposomes, the gas filled liposomes being substantially devoid of any liquid in the interior thereof. The following examples should not be construed as limiting the scope of the appended claims.

EXAMPLES

Example 1

Dipalmitoylphosphatidylcholine (1 gram) is suspended in 10 ml phosphate buffered saline containing the drug adriamycin, the suspension is heated to about 50°C, and then is swirled by hand in a round bottom flask for about 30 minutes. The heat source is removed, and the suspension is swirled for two additional hours, while allowing the suspension to cool to room temperature, to form drug containing liposomes.

The liposomes thus prepared are placed in a vessel in an apparatus similar to that shown in Figure 1, cooled to about -10°C, and are then subjected to high negative vacuum pressure. The temperature of the liposomes is then raised to about 10°C. High negative vacuum pressure is maintained for about 48 hours. After about 48 hours, nitrogen gas is gradually instilled into the chamber over a period of about 4 hours after which time the pressure is returned to ambient pressure. The resulting drug containing vacuum dried gas instilled liposomes, the gas filled liposomes being substantially devoid of any liquid in the interior thereof, are then suspended in 10 cc of phosphate buffered saline, and then stored at about 4°C for about three months.

Example 2

To test the liposomes of Example 1 ultrasonographically, a 250 mg sample of these liposomes is suspended in 300 cc of degassed phosphate buffered saline (that is, degassed under vacuum pressure). The liposomes are then scanned *in vitro* at varying time intervals with a 7.5 mHz transducer using an Acoustic Imaging Model 5200 scanner (Acoustic Imaging, Phoenix, AZ) and employing the system test software to measure dB reflectivity. The system is standardized prior to testing the liposomes with a phantom of known acoustic impedance. Good dB reflectivity of the liposomes is shown.

Example 3

Dipalmitoylphosphatidylcholine (1 gram) and the cryoprotectant trehalose (1 gram) are suspended in 10 ml phosphate buffered saline containing the drug amphotericin-B, the suspension is heated to about 50°C, and then is swirled by hand in a round bottom flask for about 30 minutes. The heat source is removed, and the suspension is swirled for about two additional hours, while allowing the suspension to cool to room temperature, to form liposomes. The liposomes thus prepared are then vacuum dried and gas instilled, substantially following the procedures shown in Example 1, resulting in drug containing vacuum dried gas instilled liposomes, the gas filled liposomes being substantially devoid of any liquid in the interior thereof. The liposomes are then suspended in 10 cc of phosphate buffered saline, and then stored at about 4°C for several weeks.

Example 4

To test the liposomes of Example 3 ultrasonographically, the procedures of Example 2 are substantially followed. Good dB reflectivity of the liposomes is shown.

Example 5

Dipalmitoylphosphatidylcholine (1 gram) is suspended in 10 ml phosphate buffered saline containing the drug cytosine arabinosine, the suspension is heated to

about 50°C, and then swirled by hand in a round bottom flask for about 30 minutes. The suspension is then subjected to 5 cycles of extrusion through an extruder device jacketed with a thermal barrel (Extruder Device™, Lipex Biomembranes, Vancouver, Canada), both with and without conventional freeze-thaw treatment prior to extrusion, while maintaining the temperature at about 50°C. The heat source is removed, and the suspension is swirled for about two additional hours, while allowing the suspension to cool to room temperature, to form liposomes.

The liposomes thus prepared are then vacuum dried and gas instilled, substantially following the procedures shown in Example 1, resulting in drug containing vacuum dried gas instilled liposomes, the gas filled liposomes being substantially devoid of any liquid in the interior thereof. The liposomes are then suspended in 10 cc of phosphate buffered saline, and then stored at about 4°C for several weeks.

Example 6

To test liposomes of Example 5 ultrasonographically, the procedures of Example 2 are substantially followed. Good dB reflectivity of the liposomes is shown.

Example 7

In order to test the stability of the drug containing liposomes of the invention, the liposomes suspension of Example 1 is passed through 2 micron polycarbonate filters in an extruder device (Extruder Device™, Lipex Biomembranes, Vancouver, Canada) five times at a pressure of about 600 psi. After extrusion treatment, the liposomes are studied ultrasonographically, as described in Example 2. Surprisingly, even after extrusion under high pressure, the liposomes of the invention substantially retain their echogenicity.

Example 8

The liposomes of Example 1 are scanned by ultrasound using transducer frequencies varying from 3 to 7.5 mHz. The results indicate that at a higher frequency

of ultrasound, the echogenicity decays more rapidly, reflecting a relatively high resonant frequency and higher energy associated with the higher frequencies.

Example 9

A patient with cancer is given an intravenous drug containing vacuum dried gas instilled liposomes, the gas filled liposomes being substantially devoid of any liquid in the interior thereof. The drug contained in the liposomes is adriamycin. As the intravenous injection is administered, the tumor is scanned ultrasonographically and via an automated software program, and the resonant frequency of the liposomes is determined. Ultrasonic energy is then focused into the tumor at the peak resonant frequency of the liposomes. The amount of ultrasonic energy is insufficient to cause any appreciable tissue heating (that is, no change in temperature greater than 2°C), however, this energy is sufficient to cause the liposomes to pop and release the adriamycin at the tumor site. In so doing, local drug delivery is accomplished using the liposomes with ultrasound.

Example 10

In a patient with a severe localized fungal infection, drug containing vacuum dried gas instilled liposomes, the gas filled liposomes being substantially devoid of any liquid in the interior thereof, are injected intravenously and ultrasound is used in a fashion substantially similar to that described in Example 9 to accomplish local drug delivery. The drug amphotericin-B, which the liposomes contain, is effectively delivered to the site of the infection.

Various modifications of the invention in addition to those shown and described herein will be apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims.

CLAIMS

What is claimed is:

1. A drug delivery system comprising a gas filled liposome prepared by a vacuum drying gas instillation method and having encapsulated therein a drug.

2. A drug delivery system of claim 1 wherein said liposomes are comprised of lipid materials selected from the group consisting of fatty acids, lysolipids, dipalmitoylphosphatidylcholine, phosphatidylcholine, phosphatidic acid, sphingomyelin, cholesterol, cholesterol hemisuccinate, tocopherol hemisuccinate, phosphatidylethanolamine, phosphatidylinositol, lysolipids, sphingomyelin, glycosphingolipids, glucolipids, glycolipids, sulphatides, lipids with ether and ester-linked fatty acids, and polymerized lipids.

3. A drug delivery system of claim 2 wherein said liposomes are comprised of dipalmitoylphosphatidylcholine.

4. A drug delivery system of claim 1 wherein said liposomes are filled with a gas selected from the group consisting of air, nitrogen, carbon dioxide, oxygen, argon, xenon, helium, and neon.

5. A drug delivery system of claim 4 wherein said liposomes are filled with nitrogen gas.

6. A drug delivery system of claim 1 wherein said liposomes are stored suspended in an aqueous medium.

7. A drug delivery system of claim 1 wherein said liposomes are stored dry.

8. A drug delivery system of claim 1 wherein said liposomes have a stability of greater than about three weeks.

9. A drug delivery system of claim 1 wherein said liposomes have a reflectivity of greater than about 2 dB.

10. A drug delivery system of claim 9 wherein said liposomes have a reflectivity of between about 2 dB and about 20 dB.

11. A drug delivery system for ultrasonic imaging comprising a gas filled liposome substantially devoid of liquid in the interior thereof and having encapsulated therein a drug.

12. A drug delivery system of claim 11 wherein said liposomes are comprised of lipid materials selected from the group consisting of fatty acids, lysolipids, dipalmitoylphosphatidylcholine, phosphatidylcholine, phosphatidic acid, sphingomyelin, cholesterol, cholesterol hemisuccinate, tocopherol hemisuccinate, phosphatidylethanolamine, phosphatidylinositol, lysolipids, sphingomyelin, glycosphingolipids, glucolipids, glycolipids, sulphatides, lipids with ether and ester-linked fatty acids, and polymerized lipids.

13. A drug delivery system of claim 12 wherein said liposomes are comprised of dipalmitoylphosphatidylcholine.

14. A drug delivery system of claim 11 wherein said liposomes are filled with a gas selected from the group consisting of air, nitrogen, carbon dioxide, oxygen, argon, xenon, helium, and neon.

15. A drug delivery system of claim 14 wherein said liposomes are filled with nitrogen gas.

16. A drug delivery system of claim 11 wherein said liposomes are stored suspended in an aqueous medium.

17. A drug delivery system of claim 11 wherein said liposomes are stored dry.

18. A drug delivery system of claim 11 wherein said liposomes have a shelf life stability of greater than about three weeks.

19. A drug delivery system of claim 11 wherein said liposomes have a reflectivity of greater than about 2 dB.

20. A drug delivery system of claim 19 wherein said liposomes have a reflectivity of between about 2 dB and about 20 dB.

21. A method for preparing a drug delivery systems comprising the following steps:

(i) placing under negative pressure liposomes having encapsulated therein a drug;

(ii) incubating said liposomes under said negative pressure for a time sufficient to remove substantially all liquid from said liposomes; and

(iii) instilling gas into said liposomes until ambient pressures are achieved.

22. A method of claim 21 further comprising allowing said liposomes to cool prior to and during step (i) to a temperature between about -10°C and about -20°C, allowing said liposomes to warm during step (ii) to a temperature between about 10°C and about 20°C, and allowing said liposomes to warm during step (iii) to ambient temperatures.

23. A method of claim 21 wherein said negative pressure is between about 700 mm Hg and about 760 mm Hg and is applied for about 24 to about 72 hours.

24. A method of claim 21 where said gas is instilled into said liposomes over a period of about 4 to about 8 hours.

25. A method of claim 21 where said gas is selected from the group consisting of air, nitrogen, carbon dioxide, oxygen, argon, xenon, neon, and helium.

26. A method of claim 25 where said gas is nitrogen.

27. A method of claim 21 further comprising, after step (iii), extruding said liposomes through at least one filter of a selected pore size.

28. A method for preparing drug delivery systems for ultrasonic imaging comprising the following steps:

(i) allowing liposomes having ncapsulated therein a drug to cool to a temperature between about -10°C and about
-20°C;

(ii) placing said liposomes under a negative pressure of between about 700 mm Hg to about 760 mm Hg;

(iii) incubating said liposomes under said negative pressure for about 24 to about 72 hours to remove substantially all liquid from said liposomes, while allowing said liposomes to warm to a temperature between about 10°C and about 20°C; and

(iv) instilling gas into said liposomes over a period of about 4 to about 8 hours until ambient pressures are achieved, while allowing said liposomes to warm to ambient temperature.

29. A method of claim 28 where said gas is selected from the group consisting of air, nitrogen, carbon dioxide, oxygen, argon, xenon, neon, and helium.

30. A method of claim 29 where said gas is nitrogen.

31. A method of claim 28 further comprising, after step (iv), extruding said liposomes through at least one filter of a selected pore size.

32. An apparatus for preparing drug delivery systems comprising:

(i) a vessel for containing liposomes having encapsulated therein a drug;

(ii) means for applying negative pressure to the vessel to draw liquid from liposomes contained in the vessel;

(iii) a conduit connecting the negative pressurizing means to the vessel, the conduit directing the flow of the liquid; and

(iv) means for introducing a gas into the liposomes contained in the vessel.

33. An apparatus of claim 32, wherein the negative pressurizing means is a vacuum pump.

34. An apparatus according to claim 32 further comprising means for cooling the liposomes contained in the vessel.

35. An apparatus according to claim 34 wherein the cooling means has means for cooling liposomes contained in the vessel to between about -10°C and about -20°C.

36. An apparatus according to claim 35 wherein the cooling means comprises an ice bath.

37. An apparatus according to claim 32 further comprising means for collecting the liquid flowing in the conduit.

38. An apparatus according to claim 37 wherein the collecting means is a trap.

39. An apparatus according to claim 38 further comprising means for cooling the trap.

40. An apparatus according to claim 38 wherein the trap comprises first and second members adapted to direct fluid flow therethrough, the members in flow communication with each other and with the conduit, the second member being helically arranged around the first member, the trap comprising a cooling means wherein the cooling means comprises an ice bath enclosing at least a portion of the first and second members.

41. An apparatus for preparing drug delivery systems comprising:

(i) a vessel for containing liposomes having encapsulated therein a drug;

(ii) means for cooling the liposomes contained in the vessel;

(iii) means for applying negative pressure to the vessel to draw liquid from liposomes contained in the vessel;

(iv) a conduit connecting the negative pressurizing means to the vessel, the conduit directing the flow of the liquid;

(v) means for collecting the liquid flowing in the conduit; and

(vi) means for introducing a gas into the liposomes contained in the vessel.

42. A method for the controlled delivery of a drug to an internal bodily region of a patient comprising:

(a) administering to the patient a drug delivery system comprising gas filled liposomes prepared by a vacuum drying gas instillation method having encapsulated therein a drug;

(b) monitoring said liposomes using ultrasound to determine the presence of said liposomes in the region; and

(c) rupturing said liposomes using ultrasound to release said drugs in said region.

43. A method according to claim 42 wherein the drug is delivered in the area of the patient's left heart.

44. A product produced by the method of claim 21.

45. A product produced by the method of claim 28.

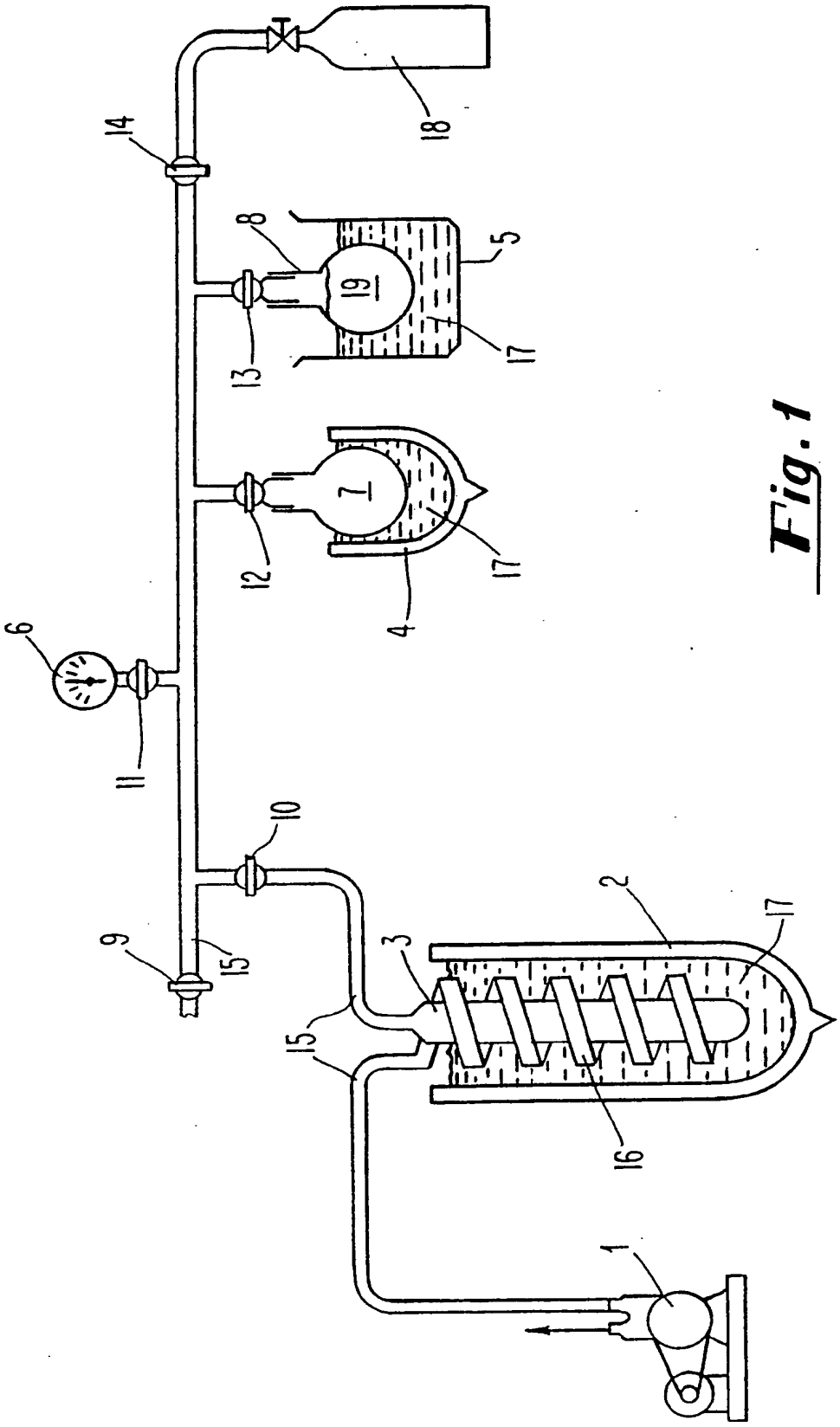

Fig. 1

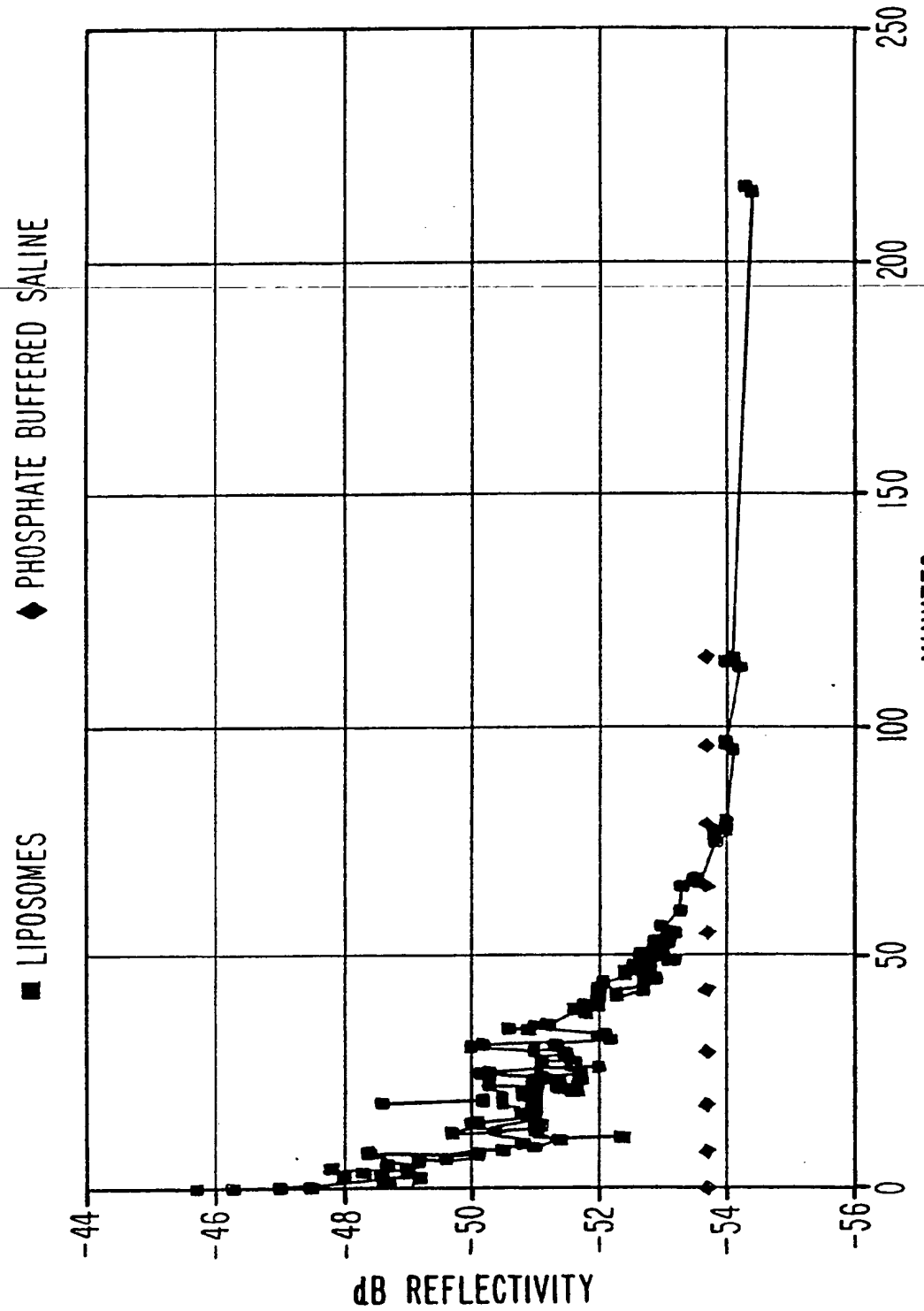
■ LIPOSOMES
◆ PHOSPHATE BUFFERED SALINE
-44
-46
-48
-50
-52
-54
-56
dB REFLECTIVITY
0
50
100
150
200
250
MINUTES

Fig. 2

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/02614

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :A61K 31/47
US CL :424/450
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 264/46; 425/5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	US,A, 4,900,540 (RYAN ET AL) 13 FEBRUARY 1990 The abstract; column 3, line 32 and claims.	1-4,6 & 44 7
X Y	US,A, 4,675,310 (CHAPMAN ET AL) 23 JUNE 1987 The abstract;	1-2,4-8 & 44 7
A	US,A, 4,657,756 (RASOR ET AL) 14 APRIL1987 The entire document.	1-45

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

*	Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A"	document defining the general state of the art which is not considered to be part of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E"	earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L"	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O"	document referring to an oral disclosure, use, exhibition or other means		
"P"	document published prior to the international filing date but later than the priority date claimed		

Date of the actual completi n f the international search 22 JULY 1992	Date of mailing of the international search report 06 OCT 1992
Name and mailing address f the ISA/ Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile N . NOT APPLICABLE	Authorized officer G.S. KISHORE Telephone No. (703) 308-2351

Form PCT/ISA/210 (second sheet)(July 1992)★

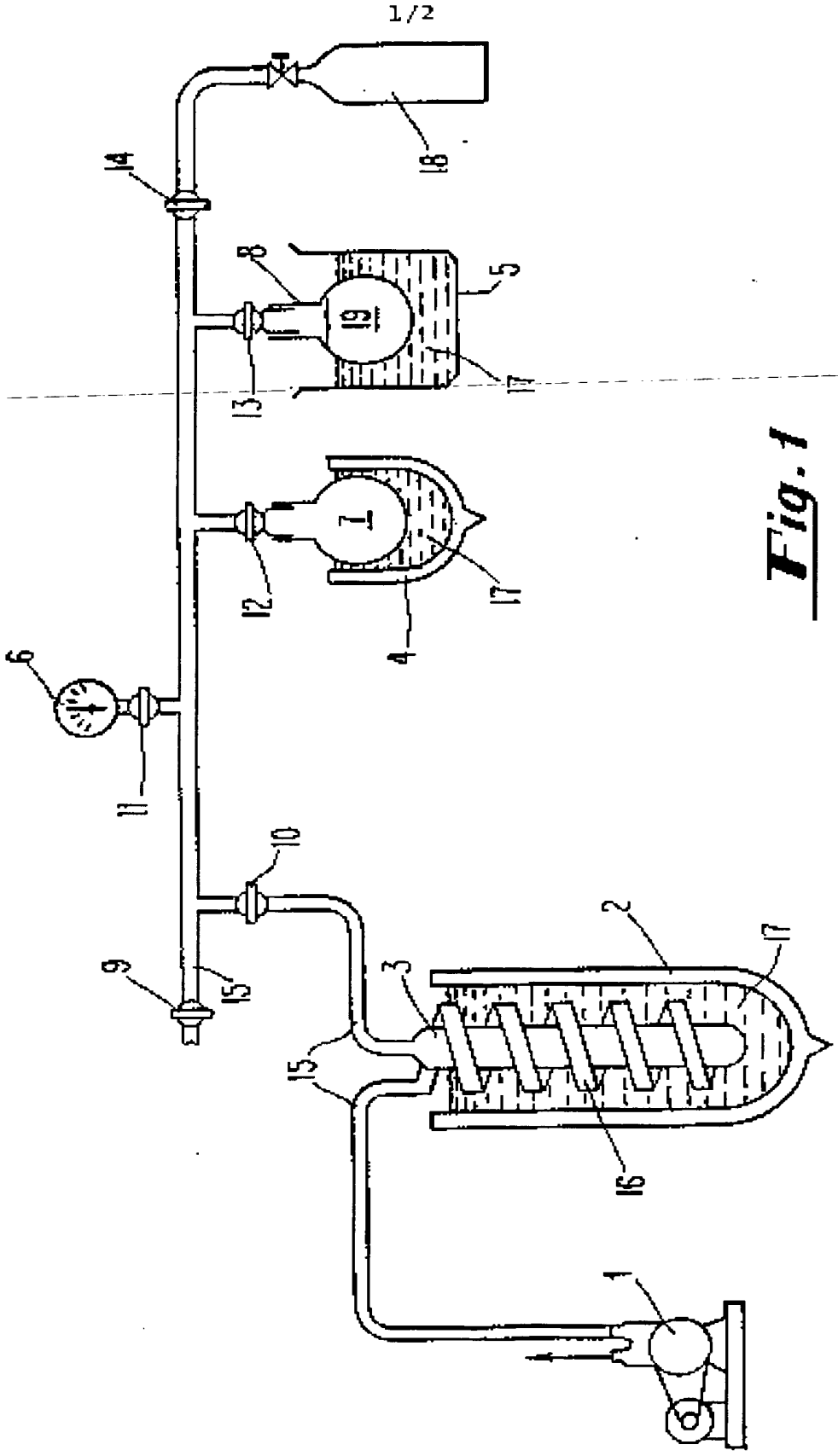

Fig. 1

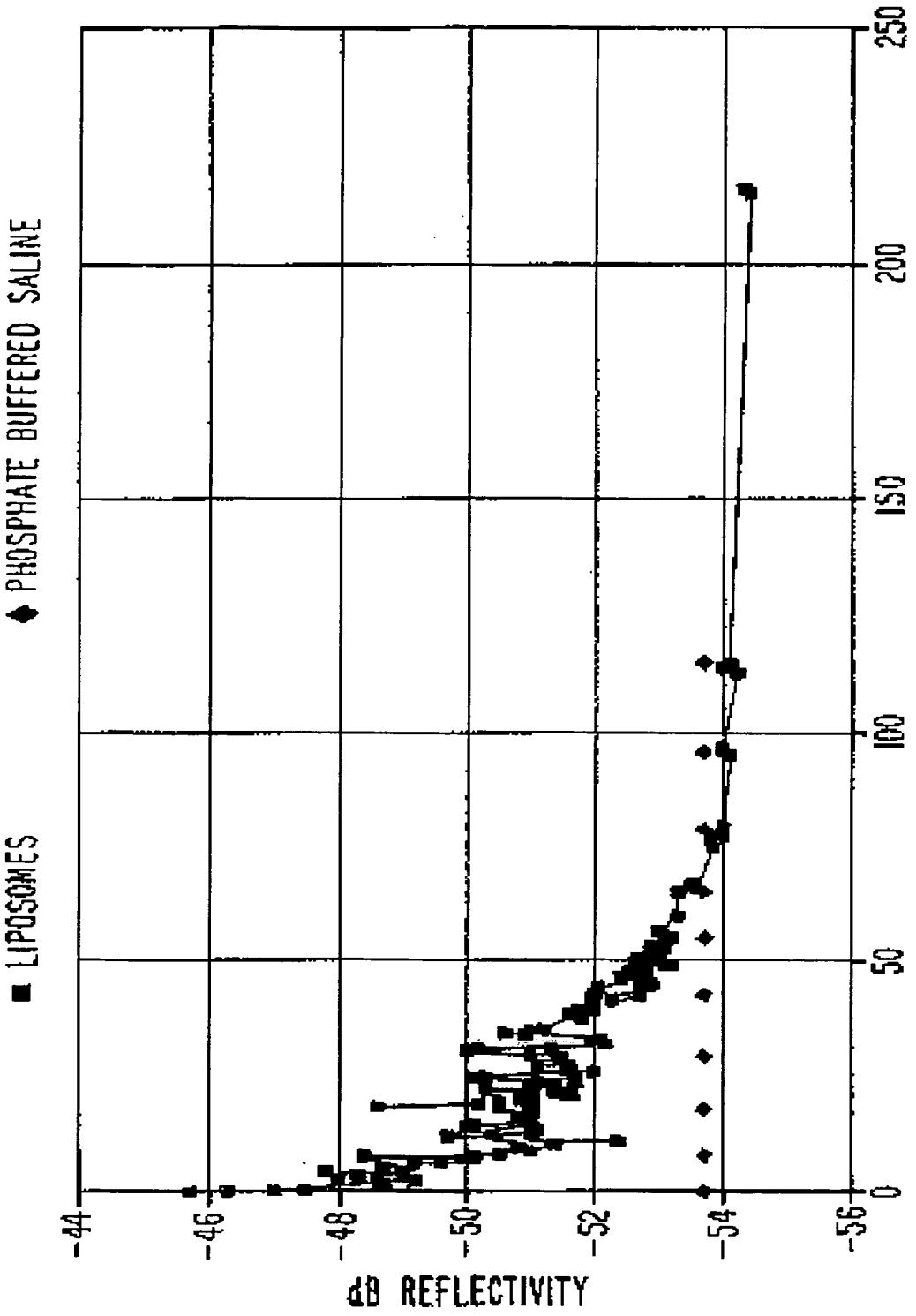

Fig. 2